

# Regulatory Roles of Novel Small Non-Coding RNAs in Antimicrobial Resistance, Biofilm Formation, Bacterial Survival, and Stress Response in Strong Biofilm-Forming *Acinetobacter baumannii*

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## BACKGROUND

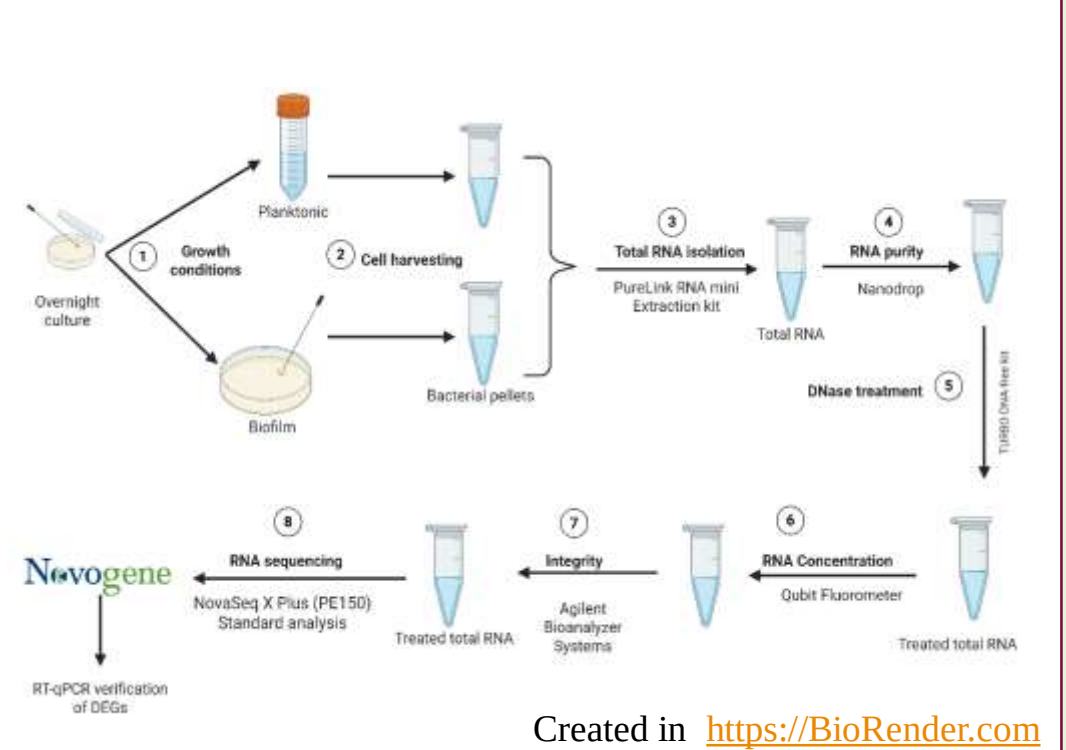
- *Acinetobacter baumannii* is a multidrug-resistant nosocomial and WHO’s critical priority pathogen for new antibiotic research. <sup>1</sup>
- It causes a wide range of human infections such as ventilator-associated pneumonia, septicemia, catheter-associated urinary tract infections, meningitis, and skin and soft tissue infections. <sup>2</sup>
- Its ability to form robust biofilm on host tissues and abiotic surfaces exacerbates the critical concerns regarding antimicrobial resistance.<sup>3</sup>
  - Biofilm inherently withstands environmental stressors, including antimicrobial agents, and therefore, they present a significant medical challenge.
  - The structure of biofilm and the physiological features of bacteria in the biofilm provide the cells with an inherent resistance to antimicrobial agents. <sup>4</sup>
- Non-coding small RNAs (sRNAs) regulate genes involved in antimicrobial resistance, biofilm formation, and virulence of various bacterial species, including *A. baumannii*. <sup>5–10</sup>

- Objective: To investigate the molecular mechanisms underlying strong biofilm formation and reduced carbapenem susceptibility in biofilm cells of *A. baumannii* ST10 clinical isolate.

## METHODS

- In our lab, 225 *A. baumannii* isolates were assessed for antimicrobial susceptibility and biofilm-forming ability.
- Among isolates originally derived from human subjects, *A. baumannii* ST10 (IC8) exhibited the strongest biofilm-forming ability.
  - ✓ It also showed broad planktonic antimicrobial susceptibility and reduced carbapenem (imipenem) susceptibility in biofilm.

### Transcriptome profiling in biofilm and planktonic cells



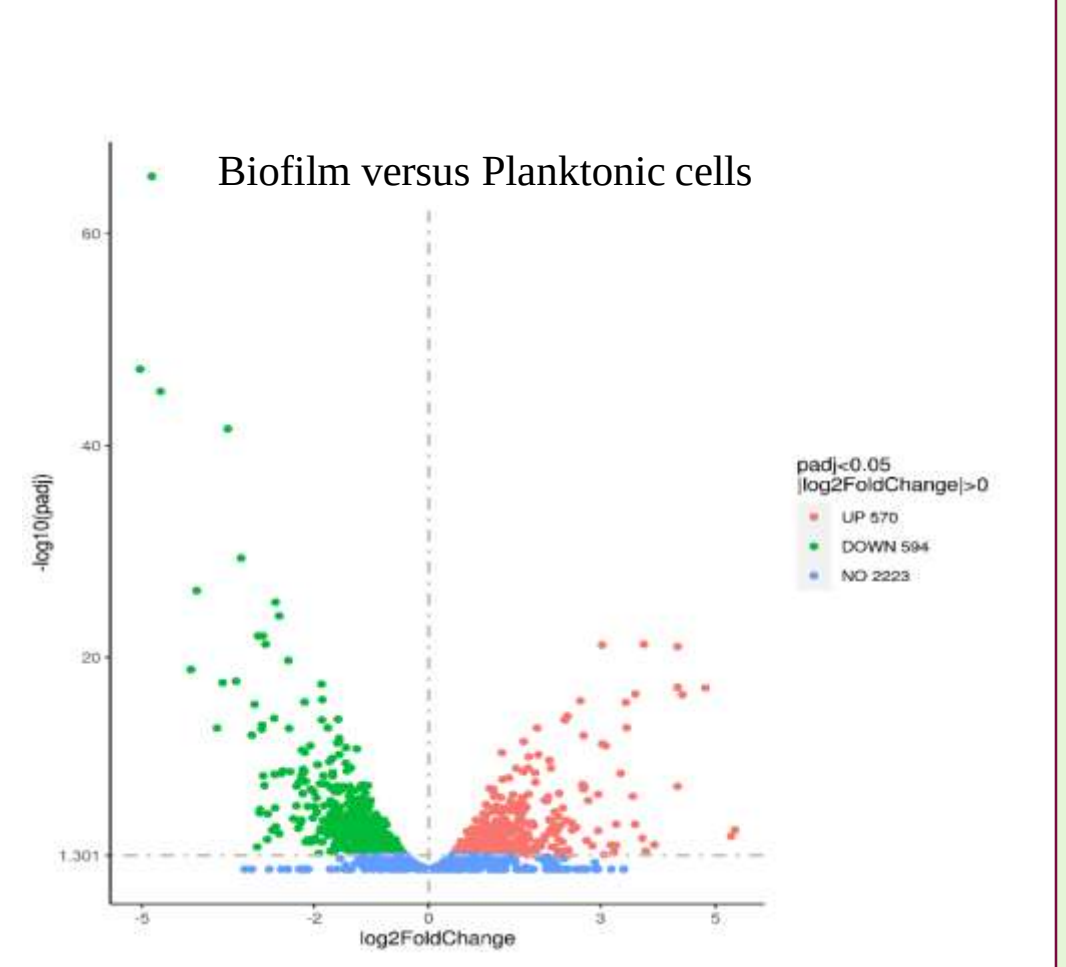
**Figure 1.** Schematic diagram showing the work-flow of the RNA samples preparation And sequencing in planktonic and biofilm cells of *A. baumannii* ST10.

**Table 1.** RNA quality assessment report for samples used in the transcriptome sequencing.

| Conditions | Sample name | Concentration (ng/µl) | Purity (Nanodrop)     |                       | RIN  |
|------------|-------------|-----------------------|-----------------------|-----------------------|------|
|            |             |                       | OD <sub>260/280</sub> | OD <sub>260/230</sub> |      |
| Biofilm    | B1          | 621                   | 2.05                  | 2.06                  | 8.50 |
|            | B2          | 623                   | 2.03                  | 2.01                  | 8.10 |
|            | B3          | 547                   | 2.00                  | 2.00                  | 8.10 |
| Planktonic | P1          | 1200                  | 2.06                  | 2.04                  | 9.20 |
|            | P2          | 1455                  | 2.04                  | 2.18                  | 9.10 |
|            | P3          | 1220                  | 2.04                  | 2.14                  | 9.40 |

## RESULTS

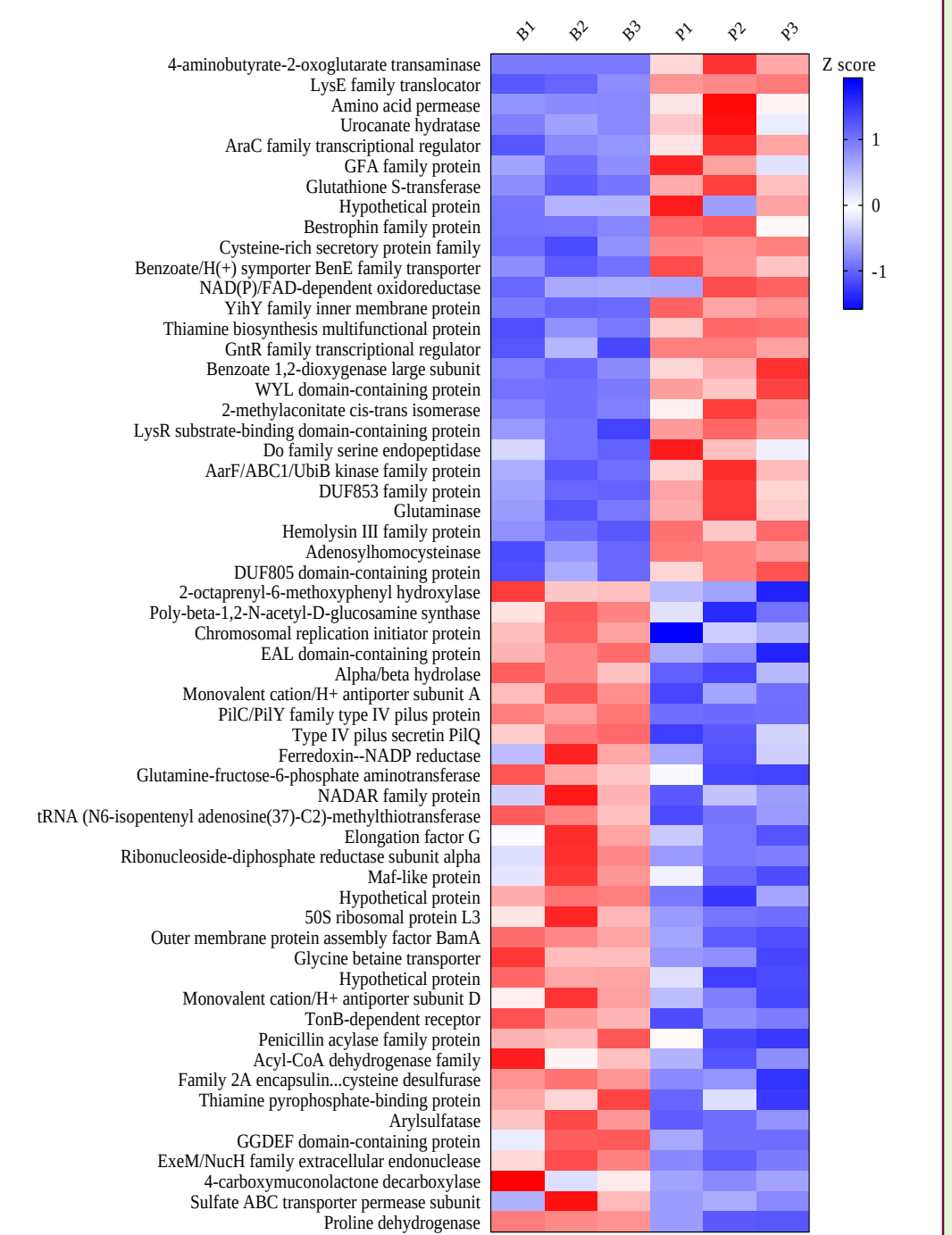
- Differentially expressed genes (DEGs)**
  - Clean bases of ~2G across all replicates in both biofilm and planktonic groups, with Q20 >98%, Q30 >96%, and an error rate of <0.01% were obtained.
  - Average mapping rates of >70% and multiple mapping rates <10% were obtained for both biofilm and planktonic cells.
  - Quantification of DEGs showed that the expressions of 1164 genes were significantly altered.



**Figure 2.** Volcano plot for differentially expressed genes between biofilm cells and planktonic cells of *A. baumannii* ST10.

### Discovery of novel sRNAs

- A total of 22 novel sRNAs were predicted from the transcriptome data.
- sRNA00007 and sRNA00048 were significantly upregulated in biofilm cells.
- In silico target prediction using IntaRNA indicated that sRNA00007 could potentially interact with a number of biofilm-associated genes.



**Figure 3.** Expression patterns of some genes that were predicted to be potential targets of sRNA00007.

### Downregulated targets of sRNA00007

- **GABA transaminase:** It is involved in GABA catabolism.
  - ✓ The GABA metabolism bypass leads to the accumulation of GABA in bacterial cells.
    - This is crucial in microbial response to environmental stresses such as desiccation, osmotic stress or acid stress. <sup>11</sup>

## RESULTS...

- **LysE translocator:** It is a group of transmembrane transport proteins involved in exporting amino acids. <sup>12</sup>
  - ✓ LysE is an essential component of bacterial growth; the absence of these exporters could result in slow growth. <sup>13</sup>
  - ✓ Biofilm cells have retarded growth compared to their planktonic counterparts. <sup>4</sup> This could be a part of a strategy to conserve energy or to alter the intracellular concentration of amino acids that are essential for biofilm maintenance and development.
- **AraC family transcriptional regulator:** It typically controls the catabolism of sugars and amino acids. <sup>14</sup>
  - ✓ Its downregulation in biofilm cells could be associated with retarded growth in biofilms.

### Upregulated targets of sRNA00007

- **TonB-dependent receptor:** It is a surface protein that plays an important role in the transportation of iron in Gram-negative bacteria. <sup>15</sup>
  - ✓ It is important for iron acquisition to promote biofilm formation.
- **ExeM/NucH family extracellular endonuclease:** It plays multiple roles, including nutrient acquisition, biofilm formation, and natural transformation of biofilm cells. <sup>16</sup>
  - ✓ It plays a central role in the modification or degradation of eDNA in biofilm.
  - ✓ It is also important in inducing biofilm dispersion.

- **GGDEF domain-containing protein:** It acts as diguanylate cyclase for the synthesis of bis-(3'-5')-cyclic dimeric guanosine monophosphate (c-di-GMP).
  - ✓ C-di-GMP is a signalling molecule (intracellular second messenger) that serves as the central biofilm regulator in Gram-negative species.
  - ✓ The accumulation of c-di-GMP promotes biofilm formation. <sup>17</sup>

## CONCLUSION AND FUTURE DIRECTIONS

- DEGs, including sRNAs in the biofilm cells, may contribute to bacterial survival under stress, enhance biofilm formation, and promote antimicrobial resistance.
- The regulatory roles of sRNA00007 and sRNA00048 in antimicrobial resistance, biofilm formation, survival, and stress response in *A. baumannii* ST10 will be further investigated.

## SELECTED REFERENCES (scan the QR code)



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